



Contents lists available at ScienceDirect

## Asian Pacific Journal of Tropical Disease

journal homepage: www.elsevier.com/locate/apjtd



Document heading

doi: 10.1016/S2222-1808(14)60643-5

© 2015 by the Asian Pacific Journal of Tropical Disease. All rights reserved.

# Detection of accessory gene regulator groups genes and cassette chromosome *mec* types among *Staphylococcus aureus* isolated from intensive care unit patients

Abdolmajid Ghasemian, Shahin Najar Peerayeh\*, Bita Bakhshi, Mohsen Mirzaee

Department of Bacteriology, Faculty of Medical Sciences, Tarbiat Modares University, Tehran, Iran

## PEER REVIEW

**Peer reviewer**

Prof. Shunhai Qu, President of Asian Pacific Tropical Medicine Press, Haikou, China.

Tel: +86-13617514235

E-mail: qu.sh@163.com

**Comments**

This is an interesting study in which the authors detected antibiotic resistance pattern, MRSA, SCCmec types and *agr* groups among isolates of *S. aureus*. Further investigations are needed to find the role of *agr* on expression of antibiotic resistance genes.

Details on Page 156

## ABSTRACT

**Objective:** To detect antibiotic resistance pattern, methicillin resistant *Staphylococcus aureus* (*S. aureus*), staphylococcal cassette chromosome *mec* (SCCmec) types and accessory gene regulator (*agr*) groups among isolates of *S. aureus*.

**Methods:** Of 78 *S. aureus* isolates, we performed antibiotic susceptibility test and then, and detected *mecA* gene, SCCmec types and *agr* specificity group genes by PCR assay.

**Results:** All the isolates were susceptible to vancomycin and linezolid. The majority of (94.2%) of methicillin resistant *S. aureus* harbored SCCmec type III. We detected *agr* group I in 45% and group II in 34.6% of the isolates. The other strains belonged to *agr* specificity groups III and IV (3.27% and 22.9%, respectively).

**Conclusions:** The majority of (45%) *S. aureus* isolates belonged to *agr* specificity group I. There was no statistical significant relationship between *agr* and antibiotic resistance and/or clinical signs.

## KEYWORDS

*Staphylococcus aureus*, Drug resistance, *Agr* groups, MRSA, SCCmec types

## 1. Introduction

*Staphylococcus aureus* (*S. aureus*) isolates cause a wide spectrum of clinical signs, ranging from mild and requiring no treatment to systemic, severe and fatal infections that occur through invasion and toxin production[1]. *S. aureus* infections occur more commonly among hospitalized and/or immunocompromised individuals[2]. Methicillin resistant *S. aureus* (MRSA) isolates can resist a laundry of antibiotics, which can make treatment of infections much more difficult.

MRSA isolates are resistant to beta-lactam antibiotics by producing penicillin-binding protein 2a with significantly

reduced affinity to beta-lactam[3]. All the staphylococcal cassette chromosome *mec* (SCCmec) types encode penicillin-binding protein 2a enzyme. Eleven SCCmec types (I–XI) have been reported, but the globally more predominant types include SCCmec I–V[4]. Moreover, MRSA isolates can be acquired from nosocomial or community origins [healthcare-associated or community-acquired (HA/CA)-MRSA][5]. CA-MRSA strains contain highly diverse and unique virulence factors.

Pathogenesis of *S. aureus* strains depends on various virulence factors which are transcriptionally controlled by a network of virulence regulators[6]. The *agr* operon plays an

\*Corresponding author: Shahin Najar Peerayeh, Department of Bacteriology, Faculty of Medical Sciences, Tarbiat Modares University, Tehran, Iran.

Tel: +98-92182883870

E-mail: najarp\_s@modares.ac.ir

Foundation Project: Supported by Tarbiat Modares University.

Article history:

Received 3 Jul 2014

Received in revised form 6 Jul, 2nd revised form 10 Jul, 3rd revised form 13 Jul 2014

Accepted 18 Jul 2014

Available online 23 Jul 2014

important role in expression of virulence genes, by encoding a specific peptide called autoinducing peptide (AIP). The *agr* operon comprises of two transcripts called RNAII and RNAIII. RNAII operon (or P2 promoter), encodes *agr* A, B, C and D. The *agr* D gene encodes AIP, which is modified by protein product of *agr* B. AIP high concentrations downregulate the expression of adhesive surface proteins, but upregulate extracellular enzymes and toxins. Furthermore, *agr* C component is a histidine kinase that acts as a sensor of AIP concentrations, and then activates *agr* A via phosphorylation mechanism. *Agr* A component is a response regulator that activates P2 or P3 promoters, either of which subsequently causes RNAII or RNAIII overexpression[7]. This two component signal transduction systems (*agr* C/A) downregulate the surface proteins and upregulate those secreted[8]. Several authors have suggested that there is a relationship between *agr* specificity group's characteristics and site of infections[3,4]. Moreover, there are reports with *agr* defective strains, because of *agr* loss during infection period[9]. The goals of this study were to determine the prevalence of MRSA, SCCmec types and *agr* specificity groups among *S. aureus* clinical isolates.

## 2. Materials and methods

### 2.1. Bacterial isolates

We collected a total of 78 *S. aureus* clinical isolates from trachea (58 isolates), blood cultures (10 isolates), lesion (6 isolates) and sputum (4 isolates) of intensive care unit patients (48 males and 30 females) from July 2012 to January 2013. In addition, the isolates were identified with catalase, slide and tube coagulases, and acid production from mannitol salt agar and DNase test.

### 2.2. Antibiotic susceptibility testing

Antimicrobial susceptibility test (AST) was conducted based on the Kirby Bauer assay (disk diffusion method), according to Clinical and Laboratory Standards Institute guidelines. We used *S. aureus* ATCC 25923 strain for quality control of antibiotic susceptibility test. Several disks were used in AST, including oxacillin (1 µg), tetracycline (30 µg), clindamycin (2 µg), erythromycin (15 µg), vancomycin (2 µg), linezolid (30 µg), trimethoprim–sulfamethoxazole (25 µg), amoxicillin (10 µg), gentamicin (10 µg) and ciprofloxacin (5 µg) (Mast Group Ltd., UK Corporation).

### 2.3. Genomic DNA extraction

Total genomic DNA was extracted via preparation of a suspension of bacterial isolates in 200 µL of tris–ethylene

diamine tetraacetic acid buffer and lysostaphin [comprising 200 µL of tris–ethylene diamine tetraacetic acid buffer and 20 µL of lysostaphin (2 µg/mL, Sigma)]. The DNA was isolated according to Straubinger method[10].

### 2.4. DNA amplification

DNA was amplified with specific primers (Table 1) to detect *mecA* gene, SCCmec types and *agr* specificity groups among the clinical isolates.

**Table 1**

Primers used for *mecA*, *agr* locus and SCCmec genes amplification.

Primer	Sequence: 3' → 5'	Product size	Reference
<i>mecA</i>	F: GTG AAG ATA TAC CAA GTG ATT R: ATG CGC TATAGATTGAAA GGA	146	11
SCCmecI	F: GCTTTAAAGACTGTCGTTACAGG R: GTTCTCTCATAGTATGACGTCC	613	11
SCCmecII	F: CGTTGAAGATGATGAAGCG R: CGAAATCAATGGTTAATGGACC	398	11
SCCmecIII	F: CCATATTGTGTACGATGCC R: CCTTACTTGTCTAACAGATCG	280	11
SCCmecIV	F: GCCTTATTGGAAGAAACCG R: CTACTCTTCTGAAAAGCGTCC	776	11
SCCmecV	F: GAACATTGTTACTTAAATGAGCG R: TGAAAGTTGTACCCTTGACACC	325	11
<i>agr</i> I	F: ATGCACATGGTGACATGC R: GTCACAAGTACTATAAGCTGCGAT	440	12
<i>agr</i> II	F: ATGCACATGGTGACATGC R: GTATTACTAATTGAAAAGTCCATAGC	572	12
<i>agr</i> III	F: ATGCACATGGTGACATGC R: CTGTTGAAAAGTCAACTAAAAGCTC	406	12
<i>agr</i> IV	F: ATGCACATGGTGACATGC R: CGATAATGCCGTAATAC CCG	588	12

F: Forward; R: Reverse.

The annealing temperature was 55 °C (30 seconds) for *mecA* gene and 51 °C (1 min) for SCCmec types, according to Zhang *et al*[11]. We carried out duplex PCR with an annealing temperature of 51 °C (1 min) for *agr* specificity groups, according to the method of Shopsin *et al*[12]. Primers for *mecA*, SCCmec types and *agr* specificity groups have been shown in Table 1. For visualization of PCR products during electrophoresis, 5 µL of each product was mixed with 1 µL of each gel red and loading buffer dyes, and was run in 1% agarose gel electrophoresis and observed by ultraviolet transilluminator.

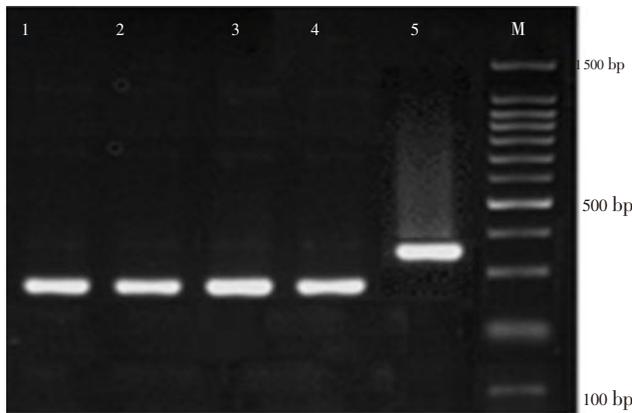
### 2.5. Statistical analysis

We used the *Chi*–square test to compare each pair of *agr* groups between methicillin–sensitive *S. aureus* (MSSA) and MRSA groups ( $P < 0.05$  was considered significant).

## 3. Results

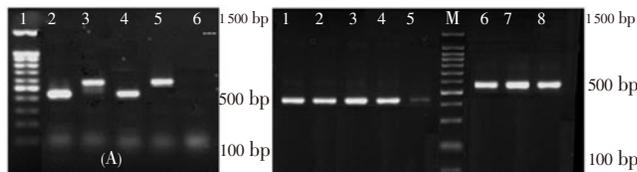
In the antibiotic susceptibility testing, 50 isolates (64%)

were resistant to amoxicillin. For the other antibiotics such as tetracycline, ciprofloxacin, gentamicin, trimethoprim–sulfamethoxazole, erythromycin and clindamycin, rate of resistance were as 21 (23%), 14 (18%), 20 (26%), 10 (13%), 23 (29.4%) and 14 (18%) isolates, respectively. Seventeen (21.7%) isolates were resistant to oxacillin and phenotypically detected as MRSA. All the studied isolates were susceptible to vancomycin and linezolid. Sixteen (94.2%) MRSA isolates harbored SCCmec type III, and one MRSA (5.8%) harbored SCCmec type V (Figures 1 and 2).



**Figure 1.** Electrophoresis of SCCmec PCR products detected (SCCmec types 3 and 5 PCR products).

Columns 1 to 4: SCCmec type III PCR products with 280 bp; Column 5: Product of SCCmec V with 325 bp; M: Marker (100 bp).



**Figure 2.** PCR products of agr specificity groups.

(A) Columns 1 and 6: Marker and control negative, respectively; Columns 2 and 4: Products of agr specificity group I (440 bp); Columns 3 and 5: agr specificity group IV (588 bp).

(B) Columns 1 to 4: agr group III (406 bp); Columns 5 to 8: Products of agr specificity group II (572 bp); M: Marker.

### 3.1. MRSA strains and agr groups

Nine (53.00%) MRSA isolates belonged to agr specificity group I, followed by agr group II in seven (41.17%) and agr specificity group IV in one (5.88%) MRSA isolate. None of them belonged to agr specificity group III (Tables 2 and 3).

**Table 2**

Prevalence of agr groups in MSSA and MRSA isolates. n (%).

Isolates	agrI	agrII	agrIII	agrIV
MSSA (n=61)	26 (42.60)	20 (32.80)	2 (3.27)	13 (21.30)
MRSA (n=17)	9 (53.00)	7 (41.17)	0	1 (5.88)
Total (n=78)	35 (57.30)	27 (44.20)	2 (3.27)	14 (22.90)

**Table 3**

Agr specificity groups' prevalence in MRSA.

Isolates (MRSA)	Clinical sample	Genus	SCCmec	agr	Antibiotic resistance
1	Bronchus	F	III	I	T, A, SXT, CD, E, CIP, GM
2	Trachea	F	III	II	T, A, SXT, CD, E, CIP, GM
3	Trachea	M	III	II	T, A
4	Lesion	M	III	II	T, A, CD, E, CIP, GM
5	Trachea	F	III	I	T, A, CD, E, CIP
6	Blood	F	III	II	T, A, CD, E, CIP
7	Trachea	M	III	I	T, A, CD, E, CIP
8	Trachea	M	III	I	T, A, CD, E, CIP
9	Lesion	M	III	II	A
10	Trachea	F	III	I	T, A, SXT, CD, E, CIP, GM
11	Trachea	M	III	I	T, A, CD, E, CIP, GM
12	Trachea	M	V	II	T, A, E, CIP
13	Trachea	M	III	I	T, A, SXT, CD, E, CIP, GM
14	Trachea	M	III	II	T, A, SXT, CD, E, CIP, GM
15	Trachea	M	III	I	T, A, SXT, CD, E, CIP, GM
16	Bronchus	M	III	I	T, A, SXT, CD, E, CIP, GM
17	Sputum	F	III	IV	T, A, SXT, CD, E, CIP, GM

F: Female; M: Male; T: Tetracycline; A: Amoxicillin; SXT: Trimethoprim–sulfamethoxazole; CD: Clindamycin; E: Erythromycin; CIP: Ciprofloxacin; GM: Gentamicin.

### 3.2. MSSA strains and agr groups

Twenty-six (42.60%) MSSA isolates belonged to agr specificity group I, and 20 (32.80%) isolates belonged to agr group II. Moreover, 2 (3.27%) and 13 (21.30%) MSSA isolates belonged to agr specificity group III and IV, respectively (Table 2). There was no significant difference regarding presence of agr groups between MRSA and MSSA strains.

## 4. Discussion

Accurate antibiotic susceptibility profile reporting is important for treatment of infections caused by *S. aureus*. In this study, we didn't determine any correlation between antibiotic resistance and origins of clinical isolates. All isolates showed susceptibility to vancomycin and linezolid, though the majority of them were resistant to amoxicillin. Seventeen (21.8%) isolates were resistant to oxacillin disk and harbored mecA gene. Two blood clinical isolates (2.5%) were resistant to methicillin and eight of these isolates (10.25%) were susceptible to this disk. Moreover, MRSA isolates with SCCmec type III were significantly more resistant to antibiotics, indicating isolates with hospital acquired infections[4]. Shittu's study results are similar to our findings, in which all of the isolates were susceptible to vancomycin and linezolid, with higher antibiotic resistance among MRSA strains[13]. In Sharma's study, all of *S. aureus* clinical isolates were susceptible to vancomycin and higher antibiotic resistance reported in MRSA isolates[14]. Similarly, Sharma et al. observed that all studied isolates were

susceptible to vancomycin and the antibiotic resistance was significantly higher in MRSA strains<sup>[15]</sup>. These results show that vancomycin and linezolid have been remained among few effective antibiotics to deal with *S. aureus* infections.

In this study, SCCmec type III was detected in 16 (98.2%) of MRSA isolates, and one isolate (5.8%) harbored SCCmec type V. These results are slightly higher than Japoni's study in south of Iran, that reported SCCmec type III in 74.3% of isolates<sup>[16]</sup>. Likewise, Fatholahzadeh reported SCCmec type III in 78% of the isolates<sup>[17]</sup>. In the study of Azimian *et al.*, SCCmec type III has been reported as the major SCCmec type<sup>[18]</sup>. In Reiter's study, all of patients with cystic fibrosis harbored SCCmec type III<sup>[19]</sup>. It seems that the clinical origin of isolates, epidemiological varieties, and the time of every study may be pivotal factors interfering in results obtained. Hospital associated MRSA isolates are resistant to a wide spectrum of antibiotics, and usually carry SCCmec type III<sup>[20]</sup>. We detected SCCmec type V in one MRSA isolate. It is believed that SCCmec type V can be acquired from community and cause severe diseases (due to Panton–Valentine leukocidin toxin production) traditionally associated to CA–MRSA<sup>[21]</sup>. There are various reports of SCCmec type V prevalence from some areas<sup>[18,19,22]</sup>. In our study, the majority of MRSA isolates (11 isolates, 65%) were from trachea, among which nine (53%) and seven (41.1%) isolates belonged to *agr* group I and II, respectively.

Various pathogenesis factors from the nosocomial strains of *S. aureus* cause the staphylococcal clinical symptoms, especially MRSA isolates containing factors that may enhance their virulence<sup>[23]</sup>. MRSA strains can affect healthy people, sometimes with high morbidity and mortality results<sup>[24]</sup>. Virulence factors of *S. aureus* are regulated by various mechanisms, such as *agr* system<sup>[25]</sup>. In this study, the majority of the isolates (54.5%) belonged to *agr* group I, similar to previous survey performed by Azimian *et al.*<sup>[26]</sup>. In addition, Indrawattana's study showed that the majority of isolates belonged to *agr* specificity group I (58.7%) and in the Ho's study, 91.6% of the clinical isolates belonged to *agr* group II<sup>[27,28]</sup>. Kahl reported that 45.7% of the clinical isolates were *agr* I positive<sup>[29]</sup>. However, Kolawole *et al.* reported that 20 of 192 isolates belonged to *agr* group II<sup>[30]</sup>. These findings indicate that *agr* group I may have an indispensable role in regulation of staphylococcal virulence. We detected no significant relationship between antibiotic resistance patterns and *agr* specificity groups. Prevalence of *agr* groups II and IV were the same (each was 18%) in this study. The *agr* group II has been more associated to respiratory infections, especially in CA–MRSA<sup>[3,31]</sup>, but our study confirmed no relationship between the origin of clinical specimens and presence of any *agr* group. The results from several studies in various areas are related potentially to some factors, such as geography, origin of specimens, and the time of conduction of studies. The rate of group III was 9% in this study, being slightly higher than Kolawole's study results that detected in 7.2% (14 of 192 isolates). There was not any significant relationship between each specific *agr* group prevalence and antibiotic susceptibility pattern of the isolates or correlation with clinical signs, similar to Indrawattana's study<sup>[27]</sup>. Further investigations are needed to find the role of *agr* on expression of antibiotic resistance genes.

The majority of MSSA and MRSA isolates belonged to *agr* group I. There was no statistical significant relationship between the *agr* groups and antibiotic resistance and/or clinical signs.

### Conflict of interest statement

We declare that we have no conflict of interest.

---

### Comments

#### Background

*S. aureus* isolates cause a wide spectrum of clinical signs, ranging from mild and requiring no treatment to systemic, severe and fatal infections that occur through invasion and toxin production. MRSA isolates can resist a laundry of antibiotics, which can make treatment of infections much more difficult.

#### Research frontiers

This study aimed to determine the prevalence of MRSA, SCCmec types and *agr* specificity groups among *S. aureus* clinical isolates.

#### Related reports

Kahl reported that 45.7% of the clinical isolates were *agr* I positive. While Kolawole *et al.* reported that 20 of 192 isolates belonged to *agr* group I.

#### Innovations & breakthroughs

This study confirmed no relationship between the origin of clinical specimens and presence of any *agr* group.

#### Applications

The study found that there was not any significant relationship between each specific *agr* group prevalence and antibiotic susceptibility pattern of the isolates or correlation with clinical signs. The results may be helpful to researchers to do further investigations.

#### Peer review

This is an interesting study in which the authors detected antibiotic resistance pattern, MRSA, SCCmec types and *agr* groups among isolates of *S. aureus*. Further investigations are needed to find the role of *agr* on expression of antibiotic resistance genes.

---

### References

- Gordon RJ, Lowy FD. Pathogenesis of methicillin-resistant *Staphylococcus aureus* infection. *Clin Infect Dis* 2008; **46**: S350–S359.
- Ghaznavi–Rad E, Nor Shamsudin M, Sekawi Z, Khoon LY, Aziz MN, Hamat RA, et al. Predominance and emergence of clones of hospital-acquired methicillin-resistant *Staphylococcus aureus* in Malaysia. *J Clin Microbiol* 2010; **48**: 867–872.
- Kim C, Milheiriço C, Gardete S, Holmes MA, Holden MT, de

- Lencastre H, et al. Properties of a novel PBP2a protein homolog from *Staphylococcus aureus* strain LGA251 and its contribution to beta-lactam-resistant phenotype. *J Biol Chem* 2012; **287**: 36854–36863.
- [4] Turlej A, Hryniewicz W, Empel J. Staphylococcal cassette chromosome *mec* (SCC*mec*) classification and typing methods: an overview. *Pol J Microbiol* 2011; **60**(2): 95–103.
- [5] Ghaznavi-Rad E, Nor Shamsudin M, Sekawi Z, van Belkum A, Neela V. A simplified multiplex PCR assay for fast and easy discrimination of globally distributed Staphylococcal cassette chromosome *mec* types in methicillin resistant *Staphylococcus aureus*. *J Med Microbiol* 2010; **59**: 1135–1139.
- [6] Robinson DA, Monk AB, Cooper JE, Feil EJ, Enright MC. Evolutionary genetics of the accessory gene regulator (*agr*) locus in *Staphylococcus aureus*. *J Bacteriol* 2005; **187**: 8312–8321.
- [7] Tsompanidou E, Sibbald MJ, Chlebowicz MA, Dreisbach A, Back JW, van Dijk JM, et al. Requirement of the *agr* locus for colony spreading of *Staphylococcus aureus*. *J Bacteriol* 2011; **193**: 1267–1272.
- [8] Balaban N, Novick RP. Translation of RNAIII, the *Staphylococcus aureus agr* regulatory RNA molecule, can be activated by a 3'-end deletion. *FEMS Microbiol Lett* 1995; **133**: 155–161.
- [9] Traber KE, Lee E, Benson S, Corrigan R, Cantera M, Shopsin B, et al. *agr* function in clinical *Staphylococcus aureus* isolates. *Microbiology* 2008; **154**: 2265–2274.
- [10] Gey A, Werckenthin C, Poppert S, Straubinger RK. Identification of pathogens in mastitis milk samples with fluorescence *in situ* hybridization. *J Vet Diagn Invest* 2013; **25**(3): 386–394.
- [11] Zhang K, McClure JA, Elsayed S, Conly JM. Novel staphylococcal cassette chromosome *mec* type, tentatively designated type VIII, harboring class A *mec* and type 4 *ccr* gene complexes in a Canadian epidemic strain of methicillin-resistant *Staphylococcus aureus*. *Antimicrob Agents Chemother* 2009; **53**(2): 531–540.
- [12] Shopsin B, Mathema B, Alcabes P, Said-Salim B, Lina G, Matsuka A, et al. Prevalence of *agr* specificity groups among *Staphylococcus aureus* strains colonizing children and their guardians. *J Clin Microbiol* 2003; **41**: 456–459.
- [13] Shittu AO, Okon K, Adesida S, Oyedara O, Witte W, Strommenger B, et al. Antibiotic resistance and molecular epidemiology of *Staphylococcus aureus* in Nigeria. *BMC Microbiol* 2011; **11**: 92.
- [14] Sharma S, Mall A. The prevalence, antibiogram and characterization of methicillin resistant *Staphylococcus aureus* among the patients from the Doon Valley hospitals. *Afr J Microbiol Res* 2011; **5**(21): 3446–3451.
- [15] Sharma NK, Garg R, Baliga S, Bhat KG. Nosocomial infections and drug susceptibility patterns in methicillin sensitive and methicillin resistant *Staphylococcus aureus*. *J Clin Diagn Res* 2013; **7**: 2178–2180.
- [16] Japoni A, Jamalidoust M, Farshad S, Ziyaeyan M, Alborzi A, Japoni S, et al. Characterization of SCC*mec* types and antibacterial susceptibility patterns of methicillin-resistant *Staphylococcus aureus* in Southern Iran. *Jpn J Infect Dis* 2011; **64**(1): 28–33.
- [17] Fatholhazadeh B, Emaneini M, Gilbert G, Udo E, Aligholi M, Modarresi MH, et al. Staphylococcal cassette chromosome *mec* (SCC*mec*) analysis and antimicrobial susceptibility patterns of methicillin-resistant *Staphylococcus aureus* (MRSA) isolates in Tehran, Iran. *Microb Drug Resist* 2008; **14**(3): 217–220.
- [18] Azimian A, Havaei SA, Fazeli H, Naderi M, Ghazvini K, Samiee SM, et al. Genetic characterization of a vancomycin-resistant *Staphylococcus aureus* isolate from the respiratory tract of a patient in a university hospital in northeastern Iran. *J Clin Microbiol* 2012; **50**(11): 3581–3585.
- [19] Reiter KC, Machado AB, Freitas AL, Barth AL. High prevalence of methicillin-resistant *Staphylococcus aureus* with SCC*mec* type III in cystic fibrosis patients in southern, Brazil. *Rev Soc Bras Med Trop* 2010; **43**(4): 377–381.
- [20] Budimir A, Deurenberg RH, Plecko V, Vink C, Kalenic S, Stobberingh EE. Molecular characterization of methicillin-resistant *Staphylococcus aureus* bloodstream isolates from Croatia. *J Antimicrob Chemother* 2006; **57**(2): 331–334.
- [21] Valsesia G, Rossi M, Bertschy S, Pfyffer GE. Emergence of SCC*mec* type IV and SCC*mec* type V methicillin-resistant *Staphylococcus aureus* containing the Panton-Valentine leukocidin genes in a large academic teaching hospital in central Switzerland: external invaders or persisting circulators? *J Clin Microbiol* 2010; **48**(3): 720–727.
- [22] Ibrahim S, Salmenlinna S, Virolainen A, Kerttula AM, Lyytikäinen O, Jägerroos H, et al. Carriage of methicillin-resistant Staphylococci and their SCC*mec* types in a long-term-care facility. *J Clin Microbiol* 2009; **47**(1): 32–37.
- [23] Gordon RJ, Lowy FD. Pathogenesis of methicillin-resistant *Staphylococcus aureus* infection. *Clin Infect Dis* 2008; **46**: S350–S359.
- [24] Miles F, Voss L, Segedin E, Anderson BJ. Review of *Staphylococcus aureus* infections requiring admission to a pediatric intensive care unit. *Arch Dis Child* 2005; **90**(12): 1274–1278.
- [25] Schlievert PM, Strandberg KL, Lin YC, Peterson ML, Leung DY. Secreted virulence factor comparison between methicillin-resistant and methicillin-sensitive *Staphylococcus aureus*, and its relevance to atopic dermatitis. *J Allergy Clin Immunol* 2010; **125**(1): 39–49.
- [26] Azimian A, Najari-Pirayeh S, Mirab-Samiee S, Naderi M. Occurrence of methicillin resistant *Staphylococcus aureus* (MRSA) among clinical samples in Tehran-Iran and its correlation with polymorphism of specific accessory gene regulator (*agr*) groups. *Braz J Microbiol* 2012; **43**(2): 779–785.
- [27] Indrawattana N, Sungkhachat O, Sookrung N, Chongsa-nguan M, Tungtrongchitr A, Voravuthikunchai SP, et al. *Staphylococcus aureus* clinical isolates: antibiotic susceptibility, molecular characteristics, and ability to form biofilm. *Biomed Res Int* 2013; doi: 10.1155/2013/314654.
- [28] Ho CM, Hsueh PR, Liu CY, Lee SY, Chiueh TS, Shyr JM, et al. Prevalence and accessory gene regulator (*agr*) analysis of vancomycin-intermediate *Staphylococcus aureus* among methicillin-resistant isolates in Taiwan—SMART program, 2003. *Eur J Clin Microbiol Infect Dis* 2010; **29**(4): 383–389.
- [29] Kahl BC, Becker K, Friedrich AW, Clasen J, Sinha B, von Eiff C, et al. *agr*-dependent bacterial interference has no impact on long-term colonization of *Staphylococcus aureus* during persistent airway infection of cystic fibrosis patients. *J Clin Microbiol* 2003; **41**(11): 5199–5201.
- [30] Kolawole DO, Adeyanju A, Schaumburg F, Akinyoola AL, Lawal OO, Amusa YB, et al. Characterization of colonizing *Staphylococcus aureus* isolated from surgical wards' patients in a Nigerian university hospital. *PLoS One* 2013; **8**(7): e68721.
- [31] Moise-Broder PA, Sakoulas G, Eliopoulos GM, Schentag JJ, Forrest A, Moellering RC Jr. Accessory gene regulator group II polymorphism in methicillin-resistant *Staphylococcus aureus* is predictive of failure of vancomycin therapy. *Clin Infect Dis* 2004; **38**(12): 1700–1705.