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First report of environmental isolation of *Cryptococcus neoformans* and other fungi from pigeon droppings in Makkah, Saudi Arabia and *in vitro* susceptibility testing

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ABSTRACT

Objective: To verify the occurrence of *Cryptococcus neoformans* (*C. neoformans*) and other fungi in samples of pigeon droppings collected from Makkah city, Saudi Arabia.

Methods: One hundred and twelve withered pigeon dropping samples were collected from 12 different districts. Using the dilution plate technique, samples were cultured on Sabouraud dextrose agar and esculin agar. Colonies were examined microscopically and *C. neoformans* identification is confirmed by India ink preparation, observation of urease activity and brown pigmentation on esculin medium. Susceptibility patterns of five yeast species and four molds against five antifungal drugs were tested using agar disk diffusion method.

Results: *C. neoformans* was recovered from 38 samples (34%). Na'am valley was recorded to be the highest contaminated site (66.7%) with *C. neoformans*, while the samples collected from Al Awaly district were considered as the lowest contaminated samples (6.7%). Also, twenty species related to sixteen genera of fungi other than *C. neoformans* were recovered from which, three yeast genera were recorded. The antifungal susceptibility testing showed that the nine tested fungal species were sensitive to Mycosat, while Fungican exerted inhibition zones of four species only. *C. neoformans* was moderately sensitive towards all tested compounds but it can resist Flucoral where no inhibition zone could be detected.

Conclusions: Our results are considered to be the first report on the environmental prevalence of *C. neoformans* in pigeon feces in Makkah, Saudi Arabia. The data indicated that pigeon droppings can be considered as a potential source of this basidiomycetous yeast in addition to other fungal species in this region.

Keywords:*Cryptococcus neoformans*

Fungi

Pigeon droppings

Susceptibility testing

Antifungal compounds

Makkah

1. Introduction

Makkah is located in a narrow valley at a height of 277 m above sea level (Figure 1a). The resident population in 2012 was roughly 7.47 million representing 26% of the total Saudi Arabian population[1]. Visitors to Makkah can reach more than 7 million every year. Makkah is characterized by the existence of a large number of free-living pigeon flocks in many sites of the city. In general, wild free-living birds (e.g. pigeons) are regarded as visible indicators of diverse and healthy environments. Yet, from a public

health standpoint, this positive view may not always be valid, since free-living birds carry a diversity of microorganisms that are pathogenic to humans[2]. Pigeon droppings have been reported as the major environmental source of *Cryptococcus neoformans* (*C. neoformans*) as well as many other pathogens in several countries[3-5]. *C. neoformans* is an encapsulated yeast-like fungus that causes cryptococcosis and its inhalation from environmental sources may cause pulmonary and neurological diseases in susceptible humans[6]. The incidence of cryptococcal infection has recently increased around the world as a result of a large increase in patients suffering from AIDS, population aging, and the expanded use of immunosuppressive drugs for cancer treatment and organ transplantation[6-8]. Since there are no available studies on the environmental distribution of *C. neoformans* in Saudi Arabia, there is an urgent need to investigate this point. The present study

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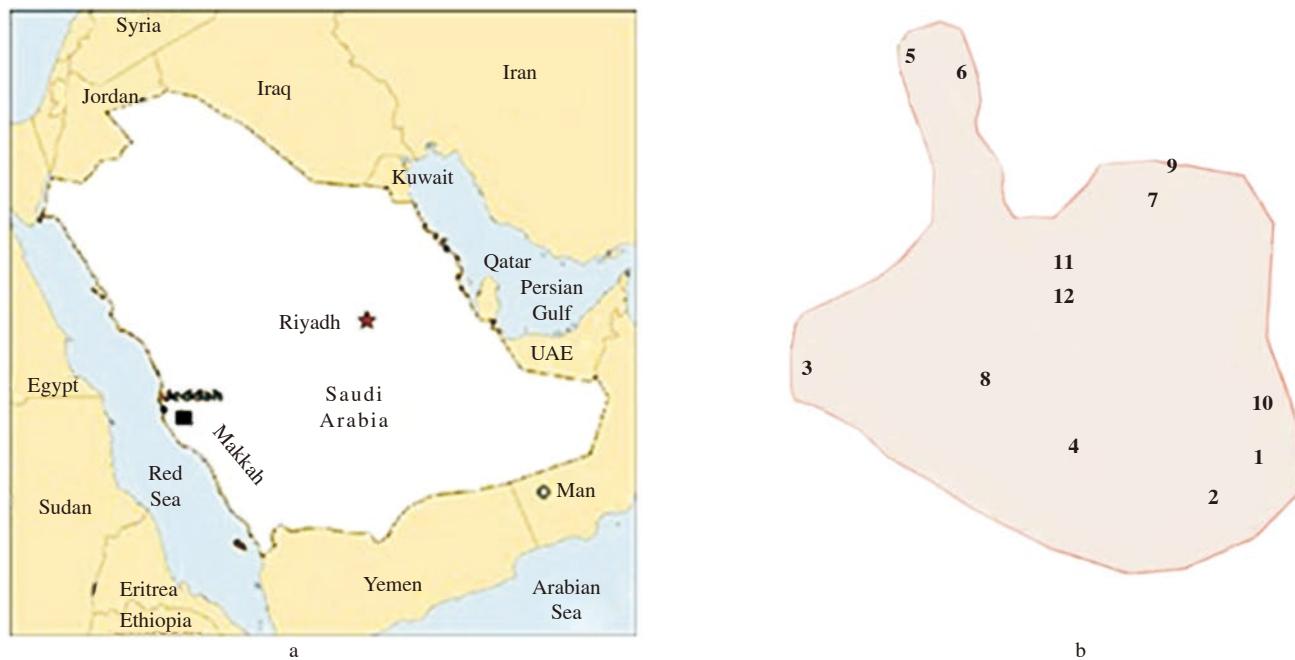


Figure 1. Saudi Arabian map showing geographical location of Makkah (a) and sampling sites in Makkah (b).

1: Al-Abedia; 2: Al-Awaly; 3: Al-Hamraa; 4: Al-Hegra; 5: Al-Nawaria; 6: Al-Omra; 7: Al-Sharayie; 8: Ashawquia; 9: Gorana; 10: Na'aman valley; 11: North Aziziya; 12: South Aziziya.

can be considered as the first step that aimed to know the extent of existence and environmental distribution of *C. neoformans* in pigeon droppings in Makkah that is considered as the highest populated city in Saudi Arabia.

2. Materials and methods

2.1. Collection of samples

Total of one hundred and twelve samples of withered pigeon droppings were collected from 12 different sites within the city (Figure 1b), over a period of six months (July to December, 2014). The samples were placed in sterile universal bottles and processed in the same day of sampling.

2.2. Isolation and identification of *C. neoformans* and other fungi

The method described by Xavier *et al.* was adapted as follows: About 10 g of withered pigeon droppings from each site were transferred to Erlenmeyer flasks containing a saline solution (0.9%) with chloramphenicol (200 mg/L), achieving 1:10 dilution (w/v) [9]. The mixture was shaken for 20 min and allowed to settle for 30 min. Aliquots of 0.5 mL supernatant were streaked on esculin agar (Oxoid, Basingstoke, UK) for isolation of *C. neoformans* and on Sabouraud dextrose agar (SDA) (Oxoid), for isolation of other fungal species [10]. The plates were then incubated at 37 °C for 10 days. Each inoculated plate was examined daily for yeast growth. Colonies with a mucous appetite and those with brown pigmentation on esculin agar (Oxoid) were selected and subcultured on SDA slants to obtain pure cultures. For identification of *C. neoformans*, morphological and biochemical tests were used. The presence of capsule on India ink preparation was considered as one of the bases of *C. neoformans* identification and confirmed by melanin synthesis on esculin agar (Oxoid), urease production on urea agar (Oxoid), and

ability to grow at 37 °C [9,11]. The other fungal species growing on SDA were identified microscopically.

2.3. Antifungal susceptibility testing

Agar disk diffusion method described by Elbanna *et al.* was mainly applied, and five antifungal compounds were purchased from different pharmacies in Makkah city to examine the antifungal susceptibility patterns of *C. neoformans* and other medically important fungal species that were recovered from pigeon feces [12]. These antifungal drugs were: Batrafen (ciclopirox-olamine), Canasten (clotrimazole), Flucoral (fluconazole), Fungican (fluconazole) and Mycosat (nystatin). The tested species were *Aspergillus flavus* (*A. flavus*), *Candida albicans* (*C. albicans*), *C. neoformans*, *Rhizopus stolonifer* (*R. stolonifer*), *Rhodotorula mucilaginosa* (*R. mucilaginosa*), *Syncephalastrum racemosum* (*S. racemosum*), *Trichoderma harzianum* (*T. harzianum*), *Trichosporon asahii* (*T. asahii*) and *Trichosporon mucoides* (*T. mucoides*). Potato dextrose agar plates were inoculated with each tested species by swabbing the fresh cultures onto the surface of agar plates. Sterilized filter paper discs (6 mm) were saturated with 100 µg/mL of each tested antifungal drug and subsequently dried at 40 °C. The dried disks were transferred to the surface of the inoculated plates in triplicates. Plates were incubated for 36 h at the optimum temperature for each fungal species and their sensitivity to the antifungal compounds was determined by measuring the diameter of inhibition zones (mm). Sterilized water without tested compounds was used as a control.

3. Results

C. neoformans was recovered from 38 out of 112 (34%) pigeon droppings in 11 out of 12 sites examined. Samples collected from site number 3 yielded negative result (Table 1). The highest rate of *C. neoformans* contamination was recorded from the samples collected

from Na'aman valley (site number 10), where 66.7% (four out of six samples) was positive. North Aziziya (site number 11) came second to Na'aman valley regarding *C. neoformans* recovery rate where it was isolated from 9 out of 16 (56.3%) samples. Al-Hegra district came third, where 50% of samples (3 out of 6) were positive (Table 1). *C. neoformans* was also recorded in four other sites (6, 1, 7 and 12) with percentage of contamination as: 40.0%, 37.5%, 33.3% and 33.3%, respectively. The occurrence of *C. neoformans* in samples collected from sites number 5, 8 and 9 were ranged between 20%-25%. On the other hand, samples collected from Al-Awaly district recorded the lowest rate of recovery, where *C. neoformans* was isolated once out of 15 samples representing 6.7% of tested samples (Table 1).

Table 1

Distribution of *C. neoformans* isolated from pigeon droppings in Makkah.

Site of isolation	Site number	Total number of samples	Positive samples	% of positive samples
Al-Abedia	1	16	6	37.5
Al-Awaly	2	15	1	6.7
Al-Hamraa	3	1	0	0.0
Al-Hegra	4	6	3	50.0
Al-Nawaria	5	4	1	25.0
Al-Omra	6	5	2	40.0
Al-Sharayie	7	9	3	33.3
Ashawquia	8	17	4	23.5
Gorana	9	5	1	20.0
Na'amman valley	10	6	4	66.7
North Aziziya	11	16	9	56.3
South Aziziya	12	12	4	33.3
Total		112	38	34.0

The isolation of other yeast and molds species from pigeon fecal droppings was shown in Table 2. Twenty species related to sixteen fungal genera were isolated within the twelve sites that were surveyed. Mucorales were represented by 8 species related to 6 genera from which *R. stolonifer*, *S. racemosum* and *Mucor hiemalis* were the most common and appeared in 16.1%, 11.6% and 10.7% of the examined samples, respectively. According to the diversity of genera, the genus *Aspergillus* ranked the second to Mucorales isolated from pigeon fecal samples and was represented by five species, namely, *A. flavus*, *Aspergillus niger* (*A. niger*), *Aspergillus parasiticus*, *Aspergillus tamarii* and *Aspergillus terreus*. The remaining genera listed in Table 2 were represented either by two or one species. *A. niger* and *R. mucilaginosa* were recorded in high frequency of occurrence and isolated from 43 and 35 pigeon fecal samples representing 38.4% and 31.3%, respectively. *A. flavus*, *R. stolonifer* and *Saccharomyces* species were encountered in moderate frequency of occurrence. The remaining fungal species were recovered from less than 15% of the samples.

Antifungal susceptibility patterns against five antifungal drugs were shown in Table 3. *C. albicans* and *T. mucoides* were sensitive to all tested antifungal compounds followed by *T. harzianum*, *C. neoformans*, *R. mucilaginosa* and *A. flavus* that were sensitive to 4 out of 5 tested compounds. *S. racemosum* and *T. asahii* were sensitive to three out of five compounds. On the other hand, *R. stolonifer* exhibited the lowest susceptibility patterns to all tested compounds. Also, it was noticed that Mycosat was found to be effective as antifungal compound against all tested isolates, followed by Canasten and Flucoral.

Table 2

Fungal genera and species (other than *C. neoformans*) isolated from pigeon droppings in Makkah.

Fungal genera and species	No. of isolation cases	Frequency of occurrence %
<i>Actinomucor elegans</i>	2	1.8R
<i>Absidia corymbifera</i>	1	0.9R
<i>A. flavus</i>	22	19.6M
<i>A. niger</i>	43	38.4H
<i>Aspergillus parasiticus</i>	13	11.6L
<i>Aspergillus tamarii</i>	3	2.7R
<i>Aspergillus terreus</i>	1	0.9R
<i>C. albicans</i>	14	12.5L
<i>Cunninghamella echinulata</i>	4	3.6R
<i>Mucor hiemalis</i>	12	10.7L
<i>Mucor racemosus</i>	10	8.9L
<i>Papulaspora candida</i>	9	8.0L
<i>Penicillium</i> spp.	7	6.3L
<i>Rhizoctonia solani</i>	4	3.6R
<i>Rhizopus oryzae</i>	2	1.8R
<i>R. stolonifer</i>	18	16.1M
<i>R. mucilaginosa</i>	35	31.3H
<i>Saccharomyces</i> spp.	18	16.1M
<i>S. racemosum</i>	13	11.6L
<i>T. harzianum</i>	1	0.9R
<i>T. asahii</i>	7	6.3L
<i>T. mucoides</i>	3	2.7R
<i>Trichosporon</i> spp.	7	6.3L
<i>Ulocladium</i> sp.	2	1.8R
Sterile mycelia dark	7	6.3L
Total	16 genera and 20 species	

H: High occurrence present in more than 30% of samples; M: Moderate occurrence present in 15% to less than 30%; L: Low occurrence present in 5% to less than 15%; R: Rare occurrence present in less than 5%.

Table 3

Sensitivity of some selected fungal species isolated from pigeon dropping samples to five pharmaceutical antifungal compounds (at concentration of 100 µg/mL).

Tested fungi	Inhibition zone in diameter (mm)				
	Batrafen	Canasten	Flucoral	Fungican	Mycosat
<i>A. flavus</i>	20	10	35	0.0	39
<i>C. albicans</i>	20	11	41	25	32
<i>C. neoformans</i>	35	28	0	27	20
<i>R. stolonifer</i>	0	0	30	0	11
<i>R. mucilaginosa</i>	27	20	60	0	43
<i>S. racemosum</i>	0	11	35	0	36
<i>T. harzianum</i>	0	15	40	10	48
<i>T. asahii</i>	0	11	24	0	35
<i>T. mucoides</i>	24	9	25	22	25

0: Resistant; < 20: Low sensitive; 20 – < 40: Moderately sensitive; ≥ 40: Highly sensitive.

4. Discussion

The current study showed that *C. neoformans* was found in 34% of 112 samples of pigeon droppings collected from 12 different sites in Makkah. Although the saprophytic distribution of *C. neoformans* in pigeon droppings was previously reported in different countries around the world such as Brazil[13], China[14], Columbia[15], Iran[16], India[9], and Italy[17], information about the environmental existence of this opportunistic fungus in Saudi Arabia is sparse. Currently there is only one published record reporting the incidence of *C. neoformans* infection in a tuberculous lymphadenitis patient in Saudi Arabia[18], therefore, the current study reports for the first time the isolation of environmental *C. neoformans* in Saudi Arabia.

It must be mentioned that, all pigeon dropping samples collected in our study were withered, based on the previous reports indicated that *C. neoformans* was not found in fresh pigeon droppings, probably because dry fecal droppings contain less bacterial flora and thus less competition for nutrients[19,20]. It is generally considered that the pigeon droppings are the main environmental source of *C. neoformans* probably due to its high content of organic material, particularly urea and creatinine[3,5,9,13,14,16,19]. Other environmental sources of the fungus include dust, chicken habitats, fruits and trees[21-23].

In this study, the relatively high rate of *C. neoformans* contamination (> 50%) in three sites (Na'am valley, Al-Hegra and north Aziziya) could be attributed to the environmental conditions such as large amount of pigeon droppings, dense populated resident and commercial areas, parks and trees, where pigeons may find appropriate locations for nesting and gathering in large numbers. On the other hand, our results showed that *C. neoformans* was recovered once from 15 pigeon fecal samples collected from Al-Awaly district. This can be explained on the basis that this district is relatively newly constructed comparing with the other districts in Makkah resulting relatively lower numbers of human and pigeon population. Thus areas inhabited by large numbers of pigeons flocks may be considered as an important ecological niche for *C. neoformans*[20].

Using the dilution plate method and SDA medium, our data showed that 20 species related to 16 genera of molds and yeasts other than *C. neoformans* were recovered from pigeon droppings. It must be mentioned that, among these 20 molds and yeasts species that were recovered, four species were of medical importance: *C. albicans*, *R. mucilaginosa*, *T. asahii* and *T. mucoides*. Costa *et al.* confirmed that urban pigeon droppings are a potential source of pathogenic yeasts and they isolate the same genera from pigeon droppings in Northeast Brazil[13]. *C. albicans* and other *Candida* spp. are commonly found in the gastrointestinal tract, oral cavity, and genital areas as harmless commensals[24]. It is worth noting that as well as being harmless commensals, *Candida* spp. are opportunistic pathogens capable of causing a wide range of superficial, localized, and/or systemic infections[25]. *Trichosporon* species are considered the second most common cause of fungaemia in patients with haematological malignant disease[25]. *Rhodotorula* species are being increasingly recognized as important human pathogens in severely immunosuppressed hosts, especially for patients with advanced HIV infection or cancer who are undergoing transplant[25-28].

In the present study, eight species related to Mucorales and five *Aspergillus* spp. in addition to unidentified *Penicillium* were recorded. Also, a number of species related to the following genera *Penicillium*, *Aspergillus*, *Mucor*, and *Rhizopus* were isolated from pigeon dropping samples collected from Isfahan[16]. Some molds recovered during the present study were previously reported to be associated with human diseases such as *Aspergillus* species[29,30]. The Mucorales group (*Mucor* spp., *Rhizopus* spp., *Absidia* spp., and *Cunninghamella* spp.) were also considered as opportunistic human pathogens[31,32]. One species of *Trichoderma* (*T. harzianum*) recorded once in our study. *T. harzianum* and *Trichoderma longibrachiatum* constitute a lethal hazard for individuals with reduced resistance including patients with leukemia, HIV positive or having transplants[33,34].

The results of antifungal susceptibility testing showed that Mycosat (nystatin) had a broad spectrum antifungal effect where it inhibited all tested fungal species. On the contrary, Fungican (fluconazole) had the narrowest antifungal effect spectrum on tested species, where five of them were resistant. *C. neoformans* showed resistance to Flucoral but was moderately sensitive to other antifungal drugs. The resistance of *C. neoformans* to Flucoral (fluconazole) was in

accordance with the finding of some other authors[35]. Varma *et al.* reported heteroresistance of *Cryptococcus gattii* to fluconazole[36]. The *in vitro* antifungal susceptibility studies of isolates of the *C. neoformans/Cryptococcus gattii* species complex revealed contradictory results[37]. The study indicated a clear correlation between antifungal susceptibilities and genotypes of the causative cryptococcosis agents. Resistance to antifungal drugs is rare among clinical isolates of *C. neoformans* but has been reported[38]. Despite there is few comparison of minimum inhibitory concentrations data between clinical and environmental isolates, the results obtained by Chowdhary *et al.* showed some differences in the patterns of susceptibility according to the origin of *C. neoformans* isolates (*i.e.* clinical or environmental)[39].

With respect to other fungal genera, our results showed that *A. flavus*, *R. stolonifer* and *S. racemosum* were resistant to Fungican (fluconazole). The isolates of *A. niger* and *A. flavus* showed varying degrees of resistance to itraconazole, ketoconazole and amphotericin B, resulting in moderate zone of inhibitions against the antifungal agents[40]. In the present study, two yeast species isolates (*C. albicans* and *T. mucoides*) were sensitive to all tested drugs, while *R. mucilaginosa* showed resistance to Fungican. Also, *T. asahii* can resist Fungican in addition to Batrafen. The resistance or sensitivity of some yeast isolates were also noticed by Sardi *et al.* who reported that 62.2% of the *C. albicans* isolates tested were susceptible to fluconazole, 15.6% to voriconazole, 91.1% to amphotericin B and 95.5% to caspofungin[41].

Our results are considered to be the first report on the environmental prevalence of *C. neoformans* in pigeon fecal samples collected from different sites of Makkah, Saudi Arabia. The data indicated that pigeon droppings can be considered as a potential source of this basidiomycetous yeast in addition to other fungal species in this region. Some of the recovered fungal species reported in this study were previously reported as pathogens associated with various types of infections in immunocompromised patients. The possibility of inhalation of the fungal spores by immunocompetent individuals, would probably lead to serious health hazards. Therefore, it is necessary to prevent the accumulations of these large flocks of pigeons for long periods of time in public areas, particularly near residential areas, public parks, food outlets and hospitals. Further studies are required to investigate the environmental distribution of *C. neoformans* and other opportunistic fungi in pigeon droppings in other populated major cities of Saudi Arabia. The *in vitro* susceptibility of nine selected fungal species with medicinal interest isolated from pigeon droppings to five conventional antifungal drugs that recommended for use in Makkah proved that tested fungal species responded variably between resistance and susceptibility.

Conflict of interest statement

We declare that we have no conflict of interest.

References

- [1] Saudi Arabian Monetary Agency. The annual report of Saudi Arabian Monetary Agency. Saudi Arabia: Saudi Arabian Monetary Agency; 2010. [Online] Available from: <http://piketty.pse.ens.fr/files/capital21c/xls/RawDataFiles/WealthReportsEtc/SovereignFunds/SaudiArabia/SAMAAnnualReportJuly2010.pdf> [Accessed on 11th April, 2015]
- [2] Abulreesh HH. Faecal shedding of antibiotic resistant *Escherichia coli* serogroups in pigeons with special reference to *E. coli* O157. *Ann Res Rev Biol* 2014; 4: 2184-91.
- [3] Dickx V, Beeckman DS, Dossche L, Tavernier P, Vanrompay D.

- Chlamydophila psittaci* in homing and feral pigeons and zoonotic transmission. *J Med Microbiol* 2010; **59**: 1348-53.
- [4] Liu Z, Ma L, Zhong Y, Wang X, Xie S. Isolation, identification and significance of *Cryptococcus neoformans* and *Candida albicans* from faecal specimen of pigeon. *Inf Technol Agric Eng* 2012; **134**: 507-12.
- [5] Rad FS. Isolation of *Cryptococcus neoformans* from pigeon excreta in Qazvin. *Life Sci J* 2013; **10**: 214-9.
- [6] Goldman JD, VoLMer ME, Luks AM. Cryptococcosis in the immunocompetent patient. *Respir Care* 2010; **55**: 1499-503.
- [7] Bratton EW, El Husseini N, Chastain CA, Lee MS, Poole C, Sturmer T, et al. Comparison and temporal trends of three groups with cryptococcosis: HIV infected; solid organ transplants and HIV-negative/non-transplants. *PLoS One* 2012; **7**: e43582.
- [8] Chae HS, Park GN, Kim SH, Jo HJ, Kim JT, Jeoung HY, et al. Rapid direct identification of *Cryptococcus neoformans* from pigeon droppings by nested PCR using CNLAC1 gene. *Poul Sci* 2012; **91**: 1983-9.
- [9] Xavier TF, Auxillia A, Kannan M, Rose AF, Kumar SR. Isolation and identification of *Cryptococcus neoformans* from pigeon droppings in Tiruchirapalli district of Tamil Nadu, South India. *Int J Curr Microbiol Appl Sci* 2013; **2**: 404-9.
- [10] Oh KS, Hwang SM. Isolation and characterization of *Cryptococcus neoformans* from environmental sources in Busan. *Mycobiology* 2005; **33**: 188-93.
- [11] Junior AMB, Santos BF, Carvalho EO, Mélo DL, Trindade RC, de Resende Stoianoff MA. Biological activity of *Cryptococcus neoformans* and *Cryptococcus gattii* from clinical and environmental isolates. *J Bras Pathol Med Lab* 2013; **49**: 160-8.
- [12] Elbanna K, Attalla K, Elbadry M, Abdeltawab A, Gamal-Eldin H, Ramadan M. Impact of floral sources and processing on the antimicrobial activities of different unifloral honeys. *Asian Pac J Trop Dis* 2014; **4**: 194-200.
- [13] Costa AK, Sidrim JJ, Cordeiro RA, Brilhante RS, Monteiro AJ, Rocha MF. Urban pigeons (*Columba livia*) as a potential source of pathogenic yeasts: a focus on antifungal susceptibility of *Cryptococcus* strains in Northeast Brazil. *Mycopathologia* 2010; **169**: 207-13.
- [14] Li AS, Pan WH, Wu SX, Hideaki T, Guo N, Shen Y, et al. Ecological surveys of *Cryptococcus* species complex in China. *Chin Med J* 2012; **125**: 511-6.
- [15] Firacative C, Torres G, Rodriguez MC, Escandón P. First environmental isolation of *Cryptococcus gattii* serotype B from Cucuta, Colombia. *Biomedica* 2011; **31**: 118-23.
- [16] Soltani M, Bayat M, Hashemi SJ, Zia M, Pestechian N. Isolation of *Cryptococcus neoformans* and other opportunistic fungi from pigeon droppings. *J Res Med Sci* 2013; **18**: 56-60.
- [17] Romeo O, Scordino F, Criseo G. Environmental isolation of *Cryptococcus gattii* serotype B VGI/MAT strains in Southern Italy. *Mycopathologia* 2011; **171**: 423-30.
- [18] Al-Tawfiq JA, Ghadour J. *Cryptococcus neoformans* abscess and osteomyelitis in an immunocompetent patient with tuberculous lymphadenitis. *Infection* 2007; **35**: 377-82.
- [19] Zarrin M, Jorfi M, Amirrajab N, Rostami M. Isolation of *Cryptococcus neoformans* from pigeon droppings in Ahwaz, Iran. *Turk J Med Sci* 2010; **40**: 313-6.
- [20] Teodoro VL, Gullo FP, Sardi Jde C, Torres EM, Fusco-Almeida AM, Mendes-Gianinni MJ. Environmental isolation, biochemical identification and antifungal drug susceptibility of *Cryptococcus* species. *Rev Soc Bras Med Trop* 2013; **46**: 759-64.
- [21] Kemoi EK, Okemo P, Bii CC. Presence of *Cryptococcus* species in domestic chicken (*Gallus gallus*) droppings and the possible risk it posed to humans in Kabigeret village, Nakuru County, Kenya. *East Afr Med J* 2012; **89**: 277-80.
- [22] Leite DP, Amadio JV, Martins ER, Simões SA, Yamamoto AC, Leal-Santos FA, et al. *Cryptococcus* spp isolated from dust microhabitats in Brazilian libraries. *J Occup Med Toxicol* 2012; **7**: 11.
- [23] Illnait-Zaragozí MT, Martínez-Machín GF, Fernández-Andreu CM, Perurena-Lancha MR, Theelen B, Beekhout T, et al. Environmental isolation and characterization of *Cryptococcus* species from living trees in Havana city, Cuba. *Mycoses* 2012; **55**: e138-44.
- [24] Abu-Elteen KH, Hamad MA. Changing epidemiology of classical and emerging human fungal infections: A review. *Jordan J Biol Sci* 2012; **5**: 215-30.
- [25] Miceli MH, Diaz JA, Lee SA. Emerging opportunistic yeast infections. *Lancet Infect Dis* 2011; **11**: 142-51.
- [26] Garcia-Suarez J, Gomez-Herruz P, Cuadros JA, Burgaleta C. Epidemiology and outcome of *Rhodotorula* infection in haematological patients. *Mycoses* 2011; **54**: 318-24.
- [27] Loss SH, Antonio AC, Roehrig C, Castro PS, Maccari JG. Meningitis and infective endocarditis caused by *Rhodotorula mucilaginosa* in an immunocompetent patient. *Rev Bras Ter Intensiva* 2011; **23**: 507-9.
- [28] Tsiodras S, Papageorgiou S, Meletiadis J, Tofas P, Pappa V, Panayiotides J, et al. *Rhodotorula mucilaginosa* associated meningitis: a subacute entity with high mortality. Case report and review. *Med Mycol Case Rep* 2014; **6**: 46-50.
- [29] Kousha M, Tadi R, Soubani OA. Pulmonary aspergillosis: a clinical review. *Eur Respir Rev* 2011; **20**: 156-74.
- [30] Thompson GR 3rd, Patterson TF. Pulmonary aspergillosis: recent advances. *Semin Respir Crit Care Med* 2011; **32**: 673-81.
- [31] Bitar D, Van Cauteren D, Lanternier F, Damnaoui E, Che D, Dromer F, et al. Increasing incidence of zygomycosis (mucormycosis), France, 1997-2006. *Emerg Infect Dis* 2009; **15**: 1395-401.
- [32] Skialda A, Pagano L, Groll A, Zimmerli S, Dupont B, Lagrou K, et al. Zygomycosis in Europe: analysis of 230 cases accrued by the registry of the European Confederation of Medical Mycology (ECMM) Working Group on Zygomycosis between 2005 and 2007. *Clin Microbiol Infect* 2011; **17**: 1859-67.
- [33] Kredics L, Antal Z, Doczi I, Manczinger L, Kevei F, Nagy E. Clinical importance of the genus *Trichoderma*. A review. *Acta Microbiol Immunol Hung* 2003; **50**: 105-17.
- [34] Trabelsi S, Hariga D, Khaled S. First case of *Trichoderma longibrachiatum* infection in a renal transplant recipient in Tunisia and review of the literatures. *Tunis Med* 2010; **88**: 52-7.
- [35] Sionov E, Chang YC, Kwon-Chang KJ. Azole heteroresistance in *Cryptococcus neoformans*: emergence of resistant clones with chromosomal disomy in the mouse brain during fluconazole treatment. *Antimicrob Agents Chemother* 2013; **57**: 5127-30.
- [36] Varma A, Kwon-Chung KJ. Heteroresistance of *Cryptococcus gattii* to fluconazole. *Antimicrob Agents Chemother* 2010; **54**: 2303-11.
- [37] Trilles L, Meyer W, Wanke B, Guarro J, Lazéra M. Correlation of antifungal susceptibility and molecular type within the *Cryptococcus neoformans/C. gattii* species complex. *Med Mycol* 2012; **50**: 328-32.
- [38] Thompson GR 3rd, Wiederhold NP, Fothergill AW, Vallor AC, Wickes BL, Patterson TF. Antifungal susceptibilities among different serotypes of *Cryptococcus gattii* and *Cryptococcus neoformans*. *Antimicrob Agents Chemother* 2009; **53**: 309-11.
- [39] Chowdhary A, Randhawa HS, Sundar G, Kathuria S, Prakash A, Khan Z, et al. *In vitro* antifungal susceptibility profiles and genotypes of 308 clinical and environmental isolates of *Cryptococcus neoformans* var. *grubii* and *C. gattii* serotype B from North-western India. *J Med Microbiol* 2011; **60**: 961-7.
- [40] Ramesh S, Kumar RS, Balaji RM. Antibiotic susceptibility of *Aspergillus* spp. isolated from contaminated food sources. *Der Pharmacia Lettre* 2013; **5**: 179-83.
- [41] Sardi JCO, Gullo FP, Pitangut NS, Fusco-Almeida AM, Mendes-Gianinni MJS. *In vitro* antifungal susceptibility of *Candida albicans* isolates from patients with chronic periodontitis and diabetes. *Clin Microbiol* 2013; **2**: 1-4.